LABORATORY REQUEST FORM FOR RABIES (HUMAN CASES)
DEPARTMENT OF MICROBIOLOGY

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<tr>
<td>Name</td>
<td>Age / Sex</td>
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<td>Ref. by: Hosp./</td>
<td>Doctor</td>
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**Type of sample:** Ante mortem: Saliva / sputum / corneal impression smear
Post mortem: Brain / spinal cord / salivary gland/ any other:

**History of exposure to animal:**
Type of Animal : Dog/cat/Cattle/Any other, specify...
Date of animal bite : Status of animal : Live / Dead
Site of bite:
Category of wound : I II III
Past H/O other animal bite :
Past H/O anti rabies vaccination :

**Clinical presentation:**
Prodormal phase : Discomfort or pain at the site / numbness or tingling in the limb /
elevated temperature/ malaise / headache / sore throat / any other specify:
Excitation phase : Restlessness / tremor / pharyngeal or laryngeal spasms / fear of water
(Hydrophobia) /convulsion on exposure to light, air, sound / intense thirst
and dehydration / any other specify:
Paralytic phase : Flaccid and limp muscle / convulsions / unconsciousness / any other Specify:

**Treatment / immunization taken:**

Sample Received Date & Time: __________  (DOCTOR'S SIGNATURE WITH NAME)
COLLECTION AND TRANSPORTATION OF SPECIMENS FOR LABORATORY DIAGNOSIS OF RABIES (HUMAN CASES)

COLLECTION OF SPECIMENS

Antemortem specimens shall include saliva/sputum, corneal smear. Postmortem specimens shall usually be brain or spinal cord. Rarely other organs included.

Saliva/Sputum:
   a) Wet a sterile cotton swab with normal saline and remove excess medium by squeezing on the sides of the vial.
   b) Swab under the tongue; make two smears each on clean-labeled glass slides.
   c) Air dry the glass slides for 10 minutes.
   d) Discard the swabs in suitable disinfectants.
   e) Wrap in paper and dispatch to the laboratory.
   f) Often due to dehydration there is little saliva in the mouth. The patient, if responsive, may be asked to cough and spit in petri dish or beaker. Mix the sputum with few ml normal saline and transfer to screw capped vial.

Corneal Smear:
   g) Retract the eyelids with thumb and one finger and press a clean marked slide against the cornea.
   h) Prepare two smears on each slide taking care to apply sufficient pressure to get the smear. Avoid exerting too much pressure as it may damage the eye.
   i) Air dry the smears for 10-15 minutes and treat with chilled acetone.

Brain:
The brain is collected at autopsy. Many times relatives do not agree for a full postmortem. In such cases Vim-Silverman needle may be used to collect a small piece of brain sample (should cover hippocampus), which is then put in 50% glycerol saline.

PRESERVATION

If possible the samples of brain and salivary glands may be sent in wide mouth leak proof containers preserved in ice. However, if the samples are to be sent long distance, these may be preserved by use of 50% glycerol saline.

LABELLING

All the specimens e.g. slides, vials must be labeled with number of specimens, name of the patient, or species of the animal, type of preservative used etc. Permanent markers should be used. The parcels should also be labeled properly.

Information to be enclosed:

(a) Hydrophobia: Name, Age, Gender, Treatment taken, Exposure to animal etc. may be enclosed.

(b) Animal: The species and breed of animal. Contact with other animal, symptoms, mode and date of death, vaccination status etc.

PACKING AND TRANSPORTATION

(a) It should preferably be wide mouth leak proof containers.

(b) Seal the mouth of the container with tape or sealing paraffin.

(c) Pack in plastic bags and put in thermocol box with sufficient ice.

(d) If sending by post, pack in sturdy wooden boxes with sufficient material (preferably absorbent cotton/saw dust/paddy husk).

(e) Transportation by courier/post.